CAP 5510: Introduction to Bioinformatics CGS 5166: Bioinformatics Tools

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Evolution and Phylogeny

Darwin: Evolution & Natural Selection

Charles Darwin's 1859 book (On the Origin of Species By Means of Natural Selection, or the Preservation of Favoured Races in the Struggle for Life) introduced the Theory of Evolution.

Struggle for existence induces a natural selection. Offspring are dissimilar from their parents (that is, variability exists), and individuals that are more fit for a given environment are selected for. In this way, over long periods of time, species evolve. Groups of organisms change over time so that descendants differ structurally and functionally from their ancestors.

Slide by Pevsner

Dominant View of Evolution

All existing organisms are derived from a common ancestor and that new species arise by splitting of a population into subpopulations that do not crossbreed.

Organization: Directed Rooted Tree; Existing species: Leaves; Common ancestor species (divergence event): Internal node; Length of an edge: Time.



At the molecular level, evolution is a process of mutation with selection.

Molecular evolution is the study of changes in genes and proteins throughout different branches of the tree of life.

Phylogeny is the inference of evolutionary relationships. Traditionally, phylogeny relied on the comparison of morphological features between organisms. Today, molecular sequence data are also used for phylogenetic analyses.

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Questions for Phylogenetic Analysis

- How many genes are related to my favorite gene?
 How related are whales, dolphins & porpoises to cows?
- Where and when did HIV or other viruses originate?
- What is the history of life on earth?
- Was the extinct quagga more like a zebra or a horse?

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Phylogenetic Trees

Molecular phylogeny uses trees to depict evolutionary relationships among organisms. These trees are based upon DNA and protein sequence data.





Tree Roots

- The root of a phylogenetic tree represents the common ancestor of the sequences. Some trees are unrooted, and thus do not specify the common ancestor.
- A tree can be rooted using an outgroup (that is, a taxon known to be distantly related from all other OTUs).

Tree nomenclature: roots



Rooted tree (specifies evolutionary path) Unrooted tree



For three operational taxonomic units (OTUs) there is one possible unrooted tree.



Any of the three edges can be selected to form a root.



Three rooted trees are possible.

Numbers of rooted and unrooted trees: 4 OTUs



For 4 OTUs there are three possible unrooted trees.

For 4 OTUs there are 15 possible rooted trees.

There is only one of these 15 trees that accurately describes the evolutionary process by which these four sequences evolved.





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Branches are unscaled...

Branches are scaled...





...OTUs are neatly aligned, and nodes reflect time

...branch lengths are proportional to number of amino acid changes

Fig. 7,8

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Examples of multifurcation: failure to resolve the branching order of some metazoans and protostomes



Rokas A. et al., Animal Evolution and the Molecular Signature of Radiations $C_{1/5/15}^{4/5/15}$ (2005), Fig. 1.

Tree nomenclature: clades

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Clade ABF (monophyletic group)



Clade group of organisms believed to have evolved from a common ancestor

Monophyletic a **group** of organisms that consists of all the descendants of a common ancestor

> Fig. 7₁8 Page 232

Tree nomenclature: clades



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time

Fig. 7₂8 Page 232



Examples of clades



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Tree nomenclature: roots

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Unrooted tree



Rooted tree (specifies evolutionary path)

Fig. 7.10 Page 234

Tree nomenclature: outgroup rooting

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Molecular Clock Hypothesis

- The molecular clock is a figurative term for a technique that uses the mutation rate of biomolecules to deduce the time in prehistory when two or more life forms diverged.
- In the 1960s, sequence data were accumulated for small, abundant proteins such as globins, cytochromes c, and fibrinopeptides.
- Some proteins appeared to evolve slowly, while others evolved rapidly.
- Linus Pauling, Emanuel Margoliash and others proposed the hypothesis of a molecular clock:
 - For every given protein, the rate of molecular evolution is approximately constant in all evolutionary lineages.

Molecular Clock Hypothesis



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Molecular Clock Hypothesis — Implications

If protein sequences evolve at constant rates, they can be used to estimate the times that sequences diverged. This is analogous to dating geological specimens by radioactive decay. Molecular evolutionary studies can be complicated by the fact that both species and genes evolve. Speciation usually occurs when a species becomes reproductively isolated. In a species tree, each internal node represents a speciation event.

Genes (and proteins) may duplicate or otherwise evolve before or after any given speciation event. The topology of a gene (or protein) based tree may differ from the topology of a species tree.

Species trees versus gene/protein trees



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Species trees versus gene/protein trees



For phylogeny, DNA can be more informative.

Some substitutions in a DNA sequence alignment can be directly observed: single nucleotide substitutions, sequential substitutions, coincidental substitutions. Additional mutational events can be inferred by analysis of ancestral sequences.

Two sequences (human and mouse) and their common ancestor: we can infer which DNA changes occurred over time

B&FG 36 Fig. 7.15

Two sequences (human and mouse) and their common ancestor: we can infer which DNA changes occurred over time

Step matrices: number of steps required to change a character

(a)

	А	С	Т	G
А	0	1	1	1
С	1	0	1	1
т	1	1	0	1
G	1	1	1	0

nucleotide step matrix

(b)

	А	С	D	Ε	F	G	Η	I	Κ	L	Μ	Ν	Ρ	Q	R	S	Т	V	W	Y	
A	0	2	1	1	2	1	2	2	2	2	2	2	1	2	2	1	1	1	2	2	
С		0	2	3	1	1	2	2	3	2	3	2	2	3	1	1	2	2	1	1	
D			0	1	2	1	1	2	2	2	3	1	2	2	2	2	2	1	3	1	
Е				0	3	1	2	2	1	2	2	2	2	1	2	2	2	1	2	2	
F					0	2	2	1	3	1	2	2	2	3	2	1	2	1	2	1	
G						0	2	2	2	2	2	2	2	2	1	1	2	1	1	2	
Η							0	2	2	1	3	1	1	1	1	2	2	2	3	1	
I								0	1	1	1	1	2	2	1	1	1	1	3	2	
Κ									0	2	1	1	2	1	1	2	1	2	2	2	
L										0	1	2	1	1	1	1	2	1	1	2	
М											0	2	2	2	1	2	1	1	2	3	
Ν												0	2	2	2	1	1	2	3	1	
Ρ													0	1	1	1	1	2	2	2	
Q														0	1	2	2	2	2	2	
R															0	1	1	2	1	2	
S																0	1	2	1	1	
Т																	0	2	2	2	
V																		0	2	2	
W																			0	2	
Y																				0	

amino acid step matrix

B&FG 3eFor amino acids, between 1 and 3 nucleotide changes areFig. 7.16required to change one residue to another.

The fundamental basis of a phylogenetic tree is a multiple sequence alignment.

(If there is a misalignment, or if a nonhomologous sequence is included in the alignment, it will still be possible to generate a tree.)

Consider the following alignment of 13 homologous globin proteins (see Fig. 3.2)

Multiple alignment of myoglobins, alpha globins, beta globins

myoglobin_kanga	MG	LSDGEWQLVLN	IWGKVETDEGGHGI	KDVLIRLFK	GHPETLE	KFDKF
myoglobin_harbo	MG	LSE <mark>G</mark> EWQLVLN	VWGKVEADLAGHG	2DVL <mark>I</mark> RLFK	GHPETLE	KFDKF
myoglobin_gray_	MG	LSDGEWHLVLN	VWGKVETDLAGHG	2EVL <mark>I</mark> RLFK	SHPETLE	KFDKF
alpha_globin_ho	MV-	LSAADKTNVKA	AWSKVGGHAGEYGA	AEALERMFI	GFPTTKT	YFPHF
alpha_globin_ka	V-	LSAADKGHVKA	IWGKVGGHAGEYAA	AEGLERTFH	SFPTTKT	YFPHF
alpha_globin_do	V-	LSPADKTNIKS	TWDKIGGHAGDYG	JEALDRTFQ	SFPTTKT	YFPHF
beta_globin_dog	MVH	LTAEEKSLVSG	LWGKVNVDEVGO	GEALGRLLI	VYPWTQR	FFDSF
beta_globin_rab	MVH	LSSEEKSAVTA	LWGKVNVEEVGO	GEALGRLLV	VYPWTQR	FFESF
beta_globin_kan	VH	LTAEEKNAITS	LWGKVAIEQTGO	JEALGRLLI	VYPWTSR	FFDHF
globin_riverlam	-PIVDSGSPAV	LSAAEKTKIRS	AWAPVYSNYETSG	JDILVKFFI	STPAAQE	FFPKF
globin_sealampr	MPIVDTGSVAP	LSAAEKTKIRS	AWAPVYSTYETSG	JDILVKFFI	STPAAQE	FFPKF
globin_soybean	VA	FTEKQDALVSS	SFEAFKANIPQYS	VFYTSILE	KAPAAKD	LFSFL
globin_insect	MKFLILALCFAAASA	LSADQISTVQA	SFDKVKGDP	JGILYAVFK	ADPSIMA	KFTQF

0 ** ***0

myoglobin kanga KHLKSEDEMKASEDLKKHGITVLTALGNILKKKGHHEAELKPLAQS---HATKHKIPVQF myoglobin harbo KHLKTEAEMKASEDLKKHGNTVLTALGGILKKKGHHDAELKPLAQS---HATKHKIPIKY myoglobin gray KHLKSEDDMRRSEDLRKHGNTVLTALGGILKKKGHHEAELKPLAQS---HATKHKIPIKY alpha globin ho -DLSHGSA----QVKAHGKKVGDALTLAVGHLDDLPGALSNLSDL---HAHKLRVDPVN alpha globin ka -DLSHGSA----QIQAHGKKIADALGQAVEHIDDLPGTLSKLSDL---HAHKLRVDPVN alpha globin do -DLSPGSA----QVKAHGKKVADALTTAVAHLDDLPGALSALSDL---HAYKLRVDPVN beta globin dog GDLSTPDAVMSNAKVKAHGKKVLNSFSDGLKNLDNLKGTFAKLSEL---HCDKLHVDPEN beta globin rab GDLSSANAVMNNPKVKAHGKKVLAAFSEGLSHLDNLKGTFAKLSEL---HCDKLHVDPEN beta globin kan GDLSNAKAVMANPKVLAHGAKVLVAFGDAIKNLDNLKGTFAKLSEL---HCDKLHVDPEN globin riverlam KGMTSADELKKSADVRWHAERIINAVNDAVASMDDTEKMSMK--DLSGKHAKSFQVDPQY globin sealampr KGLTTADQLKKSADVRWHAERIINAVNDAVASMDDTEKMSMKLRDLSGKHAKSFQVDPQY globin soybean ANPTDG----VNPKLTGHAEKLFALVRDSAGQL-KASGTVVADAALGSVHAQKAVTNPEF globin insect AG-KDLESIKGTAPFEIHANRIVGFFSKIIGELPNIEADVNTFVAS---HKPRGVTHDQ-

TTT TOO O 0 0

myoglobin_kanga	LEFISDAI	IQVIQSKHA	GNFGADAQA	AM <mark>K</mark> KALEL	FRHD	МААКҮКІ	EFGFQG
myoglobin_harbo	LEFISEAI	IHVLHSRHP.	AEFGADAQG	AMNKALEI	FRKD	IATKYKI	ELGFHG
myoglobin_gray_	LEFIS <mark>EA</mark> I	IHVLHSKHP.	AEFGADAQA	AM <mark>K</mark> KALEI	FRND	IAAKYKI	ELGFHG
alpha_globin_ho	FKLLSHCI	LSTLAVHLP	NDFTPAVHAS	SLDKFLSS	SVSTV	LTSKYR	
alpha_globin_ka	FKLLSHCI	LVTFAAHLG	DAFTPEVHAS	SLDKFLAA	VSTV	LTSKYR	
alpha_globin_do	FKLLSHCI	LVTLACHHP	TEFTPAVHAS	SLDKFFAA	VSTV	LTSKYR	
beta_globin_dog	FKLLGNVI	VCVLAHHFG	KEFTPQVQA	AY <mark>Q</mark> KVVAG	VANA	LAHKYH	
beta_globin_rab	FRLLGNVI	VIVLSHHFG	KEFTPQVQA	ay <mark>q</mark> kvvag	VANA	LAHKYH	
beta_globin_kan	FKLLG <mark>NI</mark> I	VICLAEHFG	KEFTIDTQV	AWQKLVAG	VANA	LAHKYH	
globin_riverlam	FKVL-AVI	ADTVAAG	DA(GFEKLSMO	CIILM	LRSAY-	
globin_sealampr	FKVLAAVI	ADTVAAG	DA(GFEKLMSM	ICIL	LRSAY	
globin_soybean	VVKEAI	LKTIKAAVG	DKWSDELSR	AWEVAYDE	LAAA	IKAK	
globin_insect	LNNFF	AGFVSYMKA	HTDFAGAEA	AWGATLDI	FFGM	IFSKM	
	:					:	

B&FG 3e Fig. 7.17 Daga 972

Open circles: positions that distinguish myoglobins, alpha globins, beta globins

	*******	0		••	****	0	000	٥
myoglobin_kanga	MC	GLSDGE	EWQLVLNI	WGKVET	DEGGHGK	DVLIRLF	KGHPET	LEKFDKF
myoglobin_harbo	M	GLSEGE	EWQLVLNV	WGKVEA	DLAGHGQ	DVLIRLF	KGHPET	LEKFDKF
myoglobin_gray_	MC	GLSDGE	EWHLVLNV	WGKVET	DLAGHGQ	EVLIRLF	KSHPET.	LEKFDKF
alpha_globin_ho	MV-	LSAAI	OKTNVKAA	WSKVGG	HAGEYGA	EALERMF	LGFPTT	KTYFPHF
alpha_globin_ka	V-	LSAAI	OKGHVKAI	WGKVGG	HAGEYAA	EGLERTF	HSFPTT	KTYFPHF
alpha_globin_do	V-	LSPAI	OKTNIKST	WDKIGG	HAGDYGG	EALDRTF	QSFPTT	KTYFPHF
beta_globin_dog	MVH	ILTAEE	EKSLVSGI	WGKV	NVDEVGG	EALGRLL	IVYPWT	QRFFDSF
beta_globin_rab	MVH	ILSSEE	EKSAVTAL	WGKV	NVEEVGG	EALGRLL	VVYPWT	QRFFESF
beta globin kan	VH	ILTAE	EKNAITSI	WGKV	AIEQTGG	EALGRLL	IVYPWT	SRFFDHF
globin_riverlam	-PIVDSGSPAV	/LSAAB	EKTKIRSA	WAPVYS	NYETSGV	DILVKFF	TSTPAA	QEFFPKF
globin_sealampr	MPIVDTGSVAR	LSAAF	EKTKIRSA	WAPVYS	TYETSGV	DILVKFF	TSTPAA	QEFFPKF
globin_soybean	VA	FTEK	DALVSSS	FEAFKA	NIPQYSV	VFYTSIL	EKAPAA	KDLFSFL
globin_insect	MKFLILALCFAAASA	ALSADO	QISTVQAS	FDKVKG	DPV	GILYAVE	KADPSI	MAKFTQF
				: .		:	*	* :

.. * :

B&FG 3e Fig. 7.17 D____ 171

[1] Confirm that all sequences are homologous

- [2] Adjust gap creation and extension penalties as needed to optimize the alignment
- [3] Restrict phylogenetic analysis to regions of the multiple sequence alignment for which data are available for all taxa (delete columns having incomplete data).

Constructing Evolutionary/Phylogenetic Trees

2 broad categories: Distance-based methods >Ultrametric >Additive: UPGMA Transformed Distance Neighbor-Joining Character-based >Maximum Parsimony >Maximum Likelihood > Bayesian Methods

Ultrametric

□ An ultrametric tree: decreasing internal node labels distance between two nodes is label of least common ancestor. □ An ultrametric distance matrix: Symmetric matrix such that for every i, j, k, there is tie for maximum of D(i,j), D(j,k), D(i,k)

Ultrametric: Assumptions

Molecular Clock Hypothesis, Zuckerkandl & Pauling, 1962: Accepted point mutations in amino acid sequence of a protein occurs at a constant rate.
 Varies from protein to protein
 Varies from one part of a protein to another

Ultrametric Data Sources

Lab-based methods: hybridization

- Take denatured DNA of the 2 taxa and let them hybridize. Then measure energy to separate.
- Sequence-based methods: distance

Ultrametric: Example

4/5/15

Ultrametric: Example

4/5/15

Ultrametric: Distances Computed

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Ultrametric: Distances Computed

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Ultrametric: Assumptions

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Ultrametric Data Sources

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Additive-Distance Trees

Additive distance trees are edge-weighted trees, with distance between leaf nodes are exactly equal to length of path between nodes.

Unrooted Trees on 4 Taxa

Four-Point Condition

If the true tree is as shown below, then 1. $d_{AB} + d_{CD} < d_{AC} + d_{BD}$, and 2. $d_{AB} + d_{CD} < d_{AD} + d_{BC}$

Unweighted pair-group method with arithmetic means (UPGMA)

	A	В	С
В	d _{AB}		
С	d _{AC}	d _{BC}	
D	d _{AD}	d _{BD}	d _{CD}

	AB	С
С	d _{(AB)C}	
D	d _{(AB)D}	d _{cD}

$$d_{(AB)C} = (d_{AC} + d_{BC})/2$$

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Transformed Distance Method

UPGMA makes errors when rate constancy among lineages does not hold.

Remedy: introduce an outgroup & make corrections

$$\Box \text{ Now apply UPGMA} + \left(\frac{\sum_{k=1}^{n} D_{kO}}{2} \right)$$

Start with a star topology.

Find the pair to separate such that the total length of the tree is minimized. The pair is then replaced by its arithmetic mean, and the process is repeated.

$$S_{12} = \frac{D_{12}}{2} + \frac{1}{2(n-2)} \sum_{k=3}^{n} (D_{1k} + D_{2k}) + \frac{1}{(n-2)} \sum_{3 \le i \le j \le n} D_{ij}$$

Neighbor-Joining

$$S_{12} = \frac{D_{12}}{2} + \frac{1}{2(n-2)} \sum_{k=3}^{n} (D_{1k} + D_{2k}) + \frac{1}{(n-2)} \sum_{3 \le i \le j \le n} D_{ij}$$

http://en.wikipedia.org/wiki/Neighbor_joining

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Neighbor-joining method

Constructing Evolutionary/Phylogenetic Trees

2 broad categories:

- Distance-based methods
 - > Ultrametric
 - > Additive:
 - UPGMA
 - Transformed Distance
 - Neighbor-Joining

Character-based

- > Maximum Parsimony
- Maximum Likelihood
- Bayesian Methods

Character-based Methods

- Input: characters, morphological features, sequences, etc.
- Output: phylogenetic tree that provides the history of what features changed. [Perfect Phylogeny Problem]
- □ one leaf/object, 1 edge per character, path ⇔changed traits

Example

Perfect phylogeny does not always exist.

	1	2	3	4	5
A	1	1	0	0	0
В	0	0	1	0	1
С	1	1	0	0	1
D	0	0	1	1	0
E	0	1	0	0	1

Maximum Parsimony

Minimize the total number of mutations implied by the evolutionary history

Examples of Character Data

	1	2	3	4	5			Characters/Sites							
A	1	1	0	0	0	Sequences	1	2	3	4	5	6	7	8	9
В	0	0	1	0	1	1	A	A	G	A	G	Т	Т	С	A
С	1	1	0	0	1	2	A	G	С	С	G	Т	Т	С	Т
D	0	0	1	1	0	3	A	G	A	Т	A	Т	С	С	A
E	0	1	0	0	1	4	A	G	A	G	A	Т	С	С	Т

Maximum Parsimony Method: Example

		Characters/Sites												
Sequences	1	2	3	4	5	6	7	8	9					
1	A	A	G	A	G	Т	Т	С	A					
2	A	G	С	С	G	Т	Т	С	Т					
3	A	G	A	Т	A	Т	С	С	A					
4	A	G	A	G	A	Т	С	С	Т					

Unrooted Trees on 4 Taxa

FIGURE 5.14 Three possible unrooted trees (I, II, and III) for four DNA sequences (1, 2, 3, and 4) that have been used to choose the most parsimonious tree. The possible phylogenetic relationships among the four sequences are shown in Newick format. The terminal nodes are marked by the sequence number and the nucleotide type at homologous positions in the extant species. Each dot on a branch means a substitution is inferred on that branch. Note that the nucleotides at the two internal nodes of each tree represent one possible reconstruction from among several alternatives. For example, the nucleotides at both the internal nodes of tree III(d) (bottom right) can be A instead of T. In this case, the two substitutions will be positioned on the branches leading to species 2 and 4. Alternatively, other combinations of nucleotides can be placed at the internal nodes. However, these alternatives will require three substitutions or more. The minipped pumber of pubstitutions required for site 9 is two.

	1	2	3	4	5	6	7	8	9
1	A	A	G	A	G	Т	Т	С	A
2	A	G	С	с	G	т	т	С	Т
3	A	G	A	т	A	Т	С	С	A
4	A	G	A	G	A	т	С	С	т

Inferring nucleotides on internal nodes

FIGURE 5.15 Nucleotides in six extant species (1-6) and inferred possible nucleotides in five ancestral species (7-11) according to the method of Fitch (1971). Unions are indicated by parentheses. Two different trees (a and b) are depicted. Note that the inference of an ancestral nucleotide at an internal node is dependent on the tree. Modified from Fitch (1971).

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Searching for the Maximum Parsimony Tree: Exhaustive Search

FIGURE 5.16 Exhaustive stepwise construction of all 15 possible trees for five OTUs. In step 1, we form the only possible unrooted tree for the first three OTUs (A, B, and C). In step 2, we add OTU D to each of the three branches of the tree in step 1, thereby generating three unrooted trees for four OTUs. In step 3, we add OTU E to each of the five branches of the three trees in step 2, thereby generating 15 unrooted trees. Additions of OTUs are shown as heavier lines. Modifed from Swofford et al. (1996).

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Searching for the Maximum Parsimony Tree: **Branch-&-Bound**

Probabilistic Models of Evolution

Assuming a model of substitution,

- $Pr{S_i(t+\Delta) = Y | S_i(t) = X},$
- Using this formula it is possible to compute the likelihood that data D is generated by a given phylogenetic tree T under a model of substitution. Now find the tree with the maximum likelihood.

 Time elapsed? Δ
 Prob of change along edge? Pr{S_i(t+Δ) = Y |S_i(t) = X}
 Prob of data? Product of prob for all edges

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FIGURE 5.19 Schematic representation of

Computing Maximum Likelihood Tree

(d) $L = L_{(1)} \times L_{(2)} \times L_{(3)} \times ... \times L_{(n)} = \prod_{i=1}^{n} L_{(i)}$ (d) $5/\ln L = \ln L_{(1)} + \ln L_{(2)} + \ln L_{(3)} + ... + L_{(n)} = \sum_{i=1}^{n} \ln L_{(i)}$ CAP5510 / CGS5166

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