## CAP 5510: Introduction to Bioinformatics CGS 5166: Bioinformatics Tools

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www.cis.fiu.edu/~giri/teach/BioinfS15.html

## Alternative Splicing

## Alternative Splicing

$\square$ Regular transcription: exons are spliced together in order of appearance
Alternative Splicing: Exons may be dropped or shuffled during splicing, causing alternatively spliced mRNA, giving rise to different proteins
$\square$ Short video:

- http://highered.mheducation.com/sites/9834092339/student_view0/ chapter16/animation_-_exon_shuffling.html
$\square$ Increases the set of proteins coded for by the genes in
 Human genome may very well code for over 100K protein soossipine the number of coding genes is around 30 K .


Mutually exclusive exons


Alternative $5^{\prime}$ donor sites

## Small RNA

## Small RNA

$\square$ What is non-coding RNA?

- Non-coding "genes" are transcribed, but do not code for proteins
$>$ Types of non-coding genes include
- tRNA
- rRNA
- Small RNA
- Thus, they are not translated, but function as RNA molecules

How can you find them?

- You can find them as transcripts in the total mRNA
$\square$ How long are the small RNAs?
- 50-250 nucleotides in length
- Functions of small RNA?
- Mainly regulation


## Functions of Non-Coding RNA

$\square$ tRNA and rRNA have critical specific functions
$\square$ Interference: bind to proteins to alter their function

- 6 S rRNA binds to RNA polymerase and regulates overall transcription
- tmRNA is involved in protein synthesis, including recycling of ribosomes,
- 4.5S rRNA regulates signal recognition particle (SRP), which is required for secretion of proteins
- RNase $P$ is involved in maturing tRNAs
- Often involved in regulating translation of target RNAs

Interference: bind to mRNA to "regulate" transcription \& translation

- DsrA and RprA both activate RpoS translation
$>$ This happens by base pairing to a region in leader sequence of RpoS mRNA and disrupting formation of hairpin to free up the ribosome loading site.
- OxyS inhibits RpoS translation


## Waters, Storz, Cell, 136(4):615-28, 2009

Bacteria possess numerous and diverse means of gene regulation using RNA molecules, including

- mRNA leaders that affect expression in cis,
- small RNAs that bind to proteins or
- small RNAs that bind to base pair with target RNAs, and
- CRISPR RNAs that inhibit the uptake of foreign DNA.


## Waters, Storz, Cell, 136(4):615-28, 2009



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## Copy Number Variation

## Copy-number variations (CNVs)

" "Mutation" where cells have an abnormal number of copies of one or more sections
$\square$ Most CNVs are stable and heritable
Like other genetic variations, some CNVs are associated with susceptibility or resistance to disease.

- EGFR (high) in non-small cell lung cancer
- CCL3L1 (high) lower susceptibility to HIV infection
- FCGR3B (low) high susceptibility to systemic lupus erythematosus and similar inflammatory autoimmune disorders
- Autism, schizophrenia, and idiopathic learning disability



## Horizontal Gene Transfer

## Transfer of Genetic Material

Vertical gene transfer happens during cell division

- Normal mode of inheriting genetic material by offspring
- Leads to tree phylogenies
$\square$ Horizontal gene transfer is process by which organism transfers genetic material to another organism that is not its offspring
- Common between different species of bacteria
- Can happen in eukaryotes between nuclear, mitochondrial and chloroplast
- Can happen between a eukaryotic cell and a viral or bacterial cell (during endocytosis)


## Horizontal Gene Transfer (HGT)

$\square$ HGT happens by nonhomologous recombination
$\square$ Three mechanisms:

- conjugation,
- transduction, or
- Transformation
$\square$ Bacteria can acquire genetic material from other co-habiting bacteria or viruses



## HGT in bacteria is important ...

Survival-specific genes acquired through HGT

- Antibiotic resistance genes
- Toxin resistance genes
$\square$ Environmental adaptation genes acquired through HGT
- "Symbiosis island" in Mesorhizobium loti
- "Metabolic island" in Salmonella senftenberg
- Pathogenicity islands carrying Virulence genes


## How to detect HGT?

$\square$ Main principle: Sequences new to genome will retain (for a while) signatures of donor genome and distinguished from ancestral DNA

- Sequence analysis
> Closest orthologs are in unrelated bacteria (e.g., w/ BLAST)
> Synteny
- G+C Content
- Unusual codon usage
- Other sequence signatures
> Dyad, Triad, k-mer frequencies
- Located close to direct and inverted repeats
- Located close to RNA genes
- Phylogenetic methods: Evolutionary history different from rest of genome
- Database of known insertion sequences
- Other factors include presence of special genes: soj, integrons, etc.
- Combination of above


## Disordered Proteins

## October 9, 2013

$\square$ Nobel Prize for Chemistry Awarded

- For computer models for complex chemical processes
- Martin Karplus, Michael Levitt, Arieh Warshel



## Sequence $\rightarrow$ Structure

> P00698 Chicken Lysozyme Protein Sequence MRSLLILVLC FLPLAALGKV FGRCELAAAM KRHGLDNYRG YSLGNWVCAA KFESNFNTQA TNRNTDGSTD YGILQINSRW WCNDGRTPGS RNLCNIPCSA LLSSDITASV NCAKKIVSDG NGMNAWVAWR NRCKGTDVQA WIRGCRL


## Sequence $\rightarrow$ Structure

Minimize total classical potential energy of the system
$\square$ Protein structure determined by the minimum energy configuration
$\square$ Potential energy is approximated by 6 terms

- $\sum K_{b}\left(b-b_{0}\right)^{2}+$
- $\Sigma K_{\theta}\left(\theta-\theta_{0}\right)^{2}+$
- $\sum K_{\Phi}\left(\Phi-\Phi_{0}\right)^{2}+$
- $\sum \varepsilon_{i j}\left(r_{i j}{ }^{\prime} / r_{i j}\right)^{12}-\sum 2 \varepsilon_{i j}\left(r_{i j}{ }^{\prime} / r_{i j}\right)^{6}+$
- $\Sigma Q_{i} Q_{j} / r_{i j}$

Bond Length
Bond Angle
Bond Angle
Lennard-Jones potential
Coulomb Forces
$\square$ The $r^{-12}$ term is Pauli repulsive term at short ranges due to overlapping electron orbitals and the $r^{-6}$ term is attractive long-range term (van der Waals force, or dispersion force).

- "Computer Simulation of protein folding," M. Levitt and A. Warshel, Nature, 253(5494): 694-698, 1975.


## Structure $\rightarrow$ Dynamics

In reality, what you get is not a static system, but a dynamic system.
$\square$ Understanding molecular dynamics is useful
$\square$ Challenges traditional paradigm that protein 3D structure can be fully determined by knowing its sequence, and that the minimum energy structure is a single stable structure
$\square$ Molecular dynamics (MD) is a computer simulation of physical movements of atoms and molecules in the context of N -body simulation

- "CHARMM: A Program for Macromolecular Energy, Minimization, and Dynamics Calculations," B. R. Brooks, R. E. Bruccoleri, B. D. Olafson, D. J. States, S. Swaminathan, and M. Karplus, J. Comp. Chem, 4: 187-217, 1983


## Dynamics $\rightarrow$ Function

$\square$ "Theoretical Studies of Enzymic Reactions : Dielectric, Electrostatic and Steric Stabilization of the Carbonium Ion in the Reaction of Lysozyme," A. Warshel and M. Levitt, J Mol Biol, 103: 227-249, 1976.
http://www.dnatube.com/video/1228/ tRNARibosome-Molecular-Dynamics-Simulation

## Disordered Proteins

$\square$ Protein that lacks a fixed or ordered three-dimensional structure
$\square$ Challenges traditional paradigm that protein function depends on fixed 3D structure
$\square$ Disordered proteins have regular functions
$\square$ Disorder may be associated with disease also
Different proteins have different levels of disorder

- Fully unstructured to partially structured
$\square$ Good prediction software exists
$\square$ DisProt, MobiDB, D2P2: database of such proteins [Kozlowski LP, Bujnicki JM. MetaDisorder: a meta-server for the prediction of intrinsic disorder in proteins. BMC Bioinformatics.
2012 May 24;13(1):111.]
[http://www.ideal.force.cs.is.nagoya-u.ac.jp/IDEAL]



## Transformer Proteins

## Optical Mapping

## Optical Mapping

Technology to identify location and order of restriction sites on large DNA molecules

- Enables efficient construction of accurate genome-scale restriction maps
- Used to scaffold \& validate genome assemblies (bacteria to human genome)

Nanocoding: improved technology for higher accuracy and throughput
$\square$ Technology:

- Large DNA fragments ( $\sim 100 \mathrm{~Kb}$ ) immobilized on surface
- Digested with one or more restriction enzymes
- Stained with fluorescent dye
- Imaging + Estimation of fragment lengths
- Results in ordered series of fragment lengths
$\square$ Source of errors
- Inaccurate estimates of lengths
- Missing or extra restriction sites
- Missing small fragments


## Optical Imaging

## a


b


C

| 82.86 |  | 17.51 | 21.39 | 14.16 | 33.22 | 24.45 | 6.76 |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
|  | 28.03 |  |  |  |  |  |  |

## Fixing Errors Using Optical Mapping


[Mendelowitz \& Pop. GigaScience 2014, 3:33]

## Bacterial Genome Assembly using OM

- Xavier BB, Sabirova J, Pieter M, Hernalsteens JP, de Greve H, Goossens H, Malhotra-Kumar S: Employing whole genome mapping for optimal de novo assembly of bacterial genomes. BMC Res Notes 2014, 7:484
Lin HC, Goldstein S, Mendelowitz L, Zhou S, Wetzel J, Schwartz DC, Pop M: AGORA: Assembly Guided by Optical Restriction Alignment. BMC Bioinformatics 2012, 13:189
- Nagarajan N, Read TD, Pop M: Scaffolding and validation of bacterial genome assemblies using optical restriction maps. Bioinformatics 2008, 24:1229-1235
$\square$ Zhou S, Kile A, Bechner M, Place M, Kvikstad E, Deng W, Wei J, Severin J, Runnheim R, Churas C, Forrest D, Dimalanta ET, Lamers C, Burland V, Blattner FR, Schwartz DC: Single-molecule approach to bacterial genomic comparisons via optical mapping. J Bacteriol 2004, 186:77737782.

