

BioPerl: Pairwise Sequence Alignment

```
use Bio::Tools::pSW;  
  
$factory = new Bio::Tools::pSW( '-matrix' =>  
    'blosum62.bla', '-gap' => 12, '-ext' => 2, );  
  
$factory->align_and_show($seq1, $seq2, STDOUT);
```

BioPerl: Running BLAST

```
# This program only shows how to invoke BLAST and store the result
use Bio::SeqIO;
use Bio::Tools::Run::RemoteBlast;
my $Seq_in = Bio::SeqIO->new (-file => $ARGV[0], -format => 'fasta');
my $query = $Seq_in->next_seq();
my $factory = Bio::Tools::Run::RemoteBlast->new( '-prog' => 'blastp',
    '-data' => 'swissprot', _READMETHOD => "Blast" );
my $blast_report = $factory->submit_blast($query);
my $result = $blast_report->next_result;
while( my $hit = $result->next_hit() ) {
    print "\thit name: ",
        $hit->name(), " significance: ", $hit->significance(), "\n";
}
# There are programs on the bioperl website that can help you automatically
  parse the information returned by BLAST.
```

BioPerl: Multiple Sequence Alignment

```
@params = ('ktuple' => 2, 'matrix' => 'BLOSUM');
```

```
$factory =
```

```
Bio::Tools::Run::Alignment::Clustalw->new(@params);
```

```
$aln = $factory->align(\@seq_array);
```

```
foreach $seq ( $aln->eachSeq() ) {  
    print $seq->seq(), "\n"; }
```

BioPerl: Structure

- Ability to store and manipulate structures.
- Modules: *Atom*, *Chain*, *Residue*, *Model*, *Entry*, *IO*
- *Atom*
 - `new`, `x`, `y`, `z`, `xyz`, `residue`, `element`,
- *Chain*, *Residue*
- *Entry*
 - `Add_model`, `chain`, `add_chain`, `residue`, `add_residue`,
`get_residue`, `add_atom`, `get_atoms`, `connect`,
`get_atom_by_serial`, `seqres`, ...
- *Model*

BioPerl: Structure

```
use Bio::Structure::IO;
$in = Bio::Structure::IO->new(-file => "inputfilename" , '-format' => 'pdb');
$out = Bio::Structure::IO->new(-file => ">outputfilename" , '-format' => 'pdb');
# note: we quote -format to keep older perl's from complaining.
while ( my $struc = $in->next_structure() ) {
    $out->write_structure($struc);
    print "Structure ",$struc->id," number of models: ",
        scalar $struc->model,"\n";
}
```

More Bioperl Modules

[Bioperl-1.0.2::Bio::Structure::SecStr::DSSP](#)

[bioperl-1.0.2::Bio::Structure::SecStr::STRIDE](#)

[bioperl-1.0.2::Bio::Symbol](#)

[bioperl-1.0.2::Bio::Tools](#)

[bioperl-1.0.2::Bio::Tools::Alignment](#)

[bioperl-1.0.2::Bio::Tools::Bplite](#)

[bioperl-1.0.2::Bio::Tools::Blast](#)

[bioperl-1.0.2::Bio::Tools::HMMER](#)

[bioperl-1.0.2::Bio::Tools::Prediction](#)

[bioperl-1.0.2::Bio::Tools::Run::Alignment](#)

[bioperl-1.0.2::Bio::Tools::Sim4](#)

[bioperl-1.0.2::Bio::Tools::StateMachine](#)

[bioperl-1.0.2::Bio::Tree](#)

[bioperl-1.0.2::Bio::TreeIO](#)

Cystic Fibrosis

- **Before 1985**
 - Genetic Disorder that makes people susceptible to chronic pulmonary infection
 - 1/25 Caucasians carriers, 1/2500 suffer from CF
- **1985**
 - 3 groups independently showed it is on 7th chromosome
- **1989**
 - 1480 aa, 27 exons, 23 Kb gene
- **Since 1989**
 - Many mutations and their effects
 - Gene Therapy



Total Cost
\$100 Million

Types of DNA sequence data

- EST/STS
- Single gene sequences
- Pre-finished DNA clone sequences
- Large genomic sequence tracts
- Ordering information missing in all except last two above.

Types of Maps

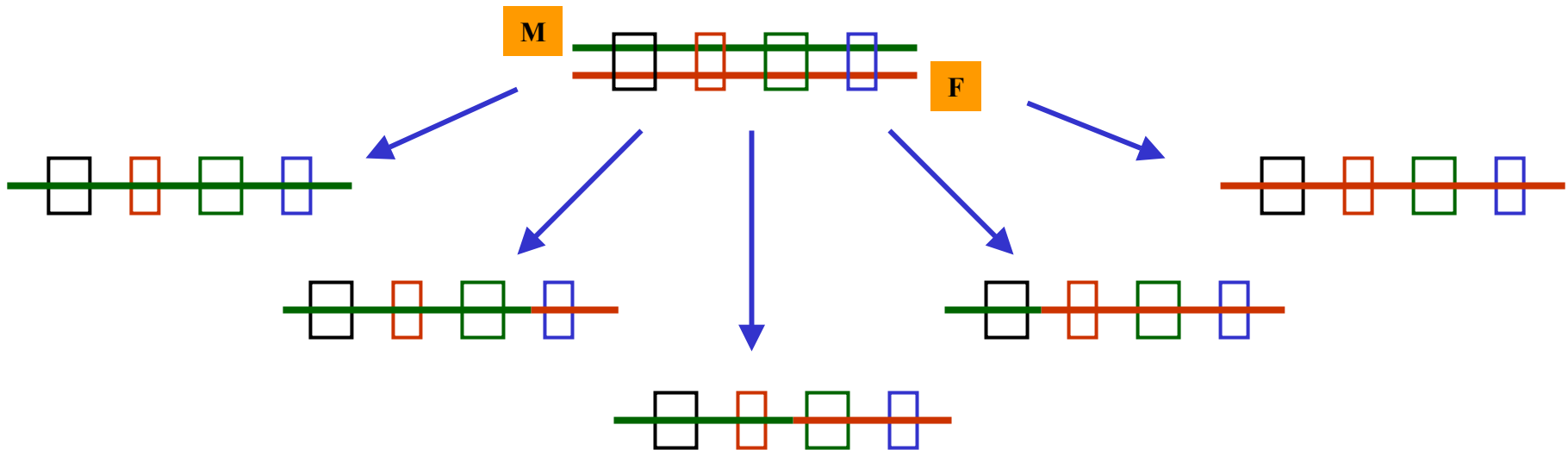
- **Cytogenetic Maps**
 - Labeled probe hybridized to chromosome, thus assigning probe to region of chromosome
- **Genetic Linkage Maps**
 - Recombination crossover events in multi-generational families used to infer relative order of markers
- **Radiation Hybrid Maps**
 - Radiation used to break chromosomes
 - PCR strength patterns used to determine proximity

Types of Maps

- EST Map
- Physical Maps
 - Precise location of various markers & genes
- Comparative Maps
 - Comparing genomes of organisms
- Integrated Maps
 - Connecting & combining different maps

Genetic Mapping

- To “approximately” locate gene in genome
- To find the order of the genes in genome
 - Sturtevant, 1913: First genetic map for 6 genes in fruit fly.



- Higher the frequency of “coinheritance” of alleles of two genes, the closer they are likely to be.

Genetic Mapping

- Analyze patterns of disease inheritance in several generations of an affected family.
- Determine which genes have alleles that are frequently **co-inherited** with the disease.
- This will help pin down **relative** location of gene on genetic map.
- Find EST that exhibits systematic difference between diseased and normal individuals and then locate gene around EST.

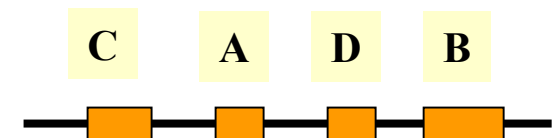
Why genetic & physical maps?

- Determine which genomic elements present in a genomic region.
- Determine their relative order.
- Determine chromosomal positions of genes.

Physical Mapping

- Determine **exact** physical location of **landmarks/markers/features** of interest.
 - STS, EST, Gene, etc.

		STS			
		A	B	C	D
Clones Fragments	1	1	0	1	0
	2	1	1	0	1
	3	1	0	0	1



Hybridization Method

	A	B	C	D	E	F	G
1	1		1				
2	1	1			1		1
3		1	1	1	1	1	1
4		1	1	1		1	1
5			1	1		1	1
6	1			1		1	1
7	1	1		1	1		1
8	1	1		1			1
9				1			

ACABEGBCDEFGBCDFG...

CAEBGCFDAGEBAGD

{CA}

{C} {A} {BEG}

CA {BEG} {CDF}

CAE {BG} {CDF} {AG}

CAE {BG} C {DF} {AG} {BE}

CAE {BG} CFD {AG} BE

Partial Digest Problem

- A restriction enzyme cuts at restriction sites.
- It produces fragments of specific lengths.



$$X = \{7, 47, 137, 177\}$$

$$\Delta X = \{7, 40, 47, 90, 100, 130, 137, 140, 170, 177, 230, 270, 277\}$$

Partial Digest Problem: Given ΔX , find X .

Double Digest Problem

- Input: 2 Restriction Enzymes A & B
 - Fragments(A)
 - Fragments(B)
 - Fragments(A & B)
- Output: Reconstruction
 - NP-Complete [[Goldstein & Waterman](#)]
 - Interesting Transformation [[Schmitt & Waterman](#)]
 - Generating all solutions [[Pevzner](#)]

Errors in practice in Physical Mapping

- False Positives
- False Negatives
 - Improper hybridization conditions
- Chimeric Clones
 - 2 clones may join together even though they are not “adjacent”
- Contamination from Host DNA